



ATTORNEY'S DOCKET NUMBER

01/22288

U.S. APPLICATION NO. (IF KNOWN), SEE 37 CFR

09/889737

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING
A FILING UNDER 35 U.S.C., 371

INTERNATIONAL APPLICATION NO.
PCT/IL00/00046

INTERNATIONAL FILING DATE
January 24, 2000

PRIORITY DATE CLAIMED
February 3, 1999

TITLE OF INVENTION

TRANSGENIC PLANTS

APPLICANT(S) FOR DO/EO/US


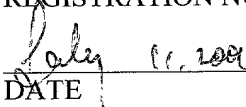
1) Jonathan GRESSEL

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(3)(2)
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(3)(2).
7. ☐ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)
 - a. ☒ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19(35 U.S.C. 371(c)(3)).
10. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
11. ☐ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 13 to 18 below concern document(s) of information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A FIRST preliminary amendment.
A SECOND or SUBSEQUENT preliminary amendment.
16. ☐ A substitute specification.
17. ☐ A change of power of attorney and/or address letter.
18. ☐ Certificate of Mailing by Express Mail
19. ☐ Other items or information:

U.S. APPLICATION NO. 09/889737 INTERNATIONAL APPLICATION NO. <div style="text-align: center;">PCT/IL00/00046</div>		ATTORNEY'S DOCKET NUMBER <div style="text-align: center;">01/22288</div>	
20. The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492(a) (1) - (5)): <input type="checkbox"/> Search Report has been prepared by the EPO or JPO \$ 860 <input checked="" type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1,482) \$ 690 <input type="checkbox"/> No international preliminary examination fee paid to USPTO (37 CFR 1,482) but international search fee paid to USPTO (37 CFR 1,445(a)(2)) \$ 710 <input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 1000 <input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$ 100 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT =</div>		CALCULATIONS PTO USE ONLY <div style="text-align: right;">\$690.00</div>	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e))		<div style="text-align: right;">\$0.00</div>	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	12 - 20 =		x \$ 18
Independent claims	6 - 3 =	3	x \$ 80
Multiple Dependent Claims (check if applicable)		<input type="checkbox"/>	\$0.00
TOTAL OF ABOVE CALCULATIONS =		\$930.00	
Reduction of 1/2 for filing by small entity, if applicable.		<input checked="" type="checkbox"/>	\$465.00
SUBTOTAL =		\$465.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f))		<div style="text-align: right;">\$0.00</div>	
TOTAL NATIONAL FEE =		\$465.00	
Fee for recording the enclosed assignment (37 CFR 1.2(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable)		<input checked="" type="checkbox"/>	\$40.00
TOTAL FEES ENCLOSED =		\$505.00	
		Amount to be refunded:	\$
		charged	\$
<input type="checkbox"/> A check in the amount of \$ _____ to cover the above fees is enclosed. <input checked="" type="checkbox"/> Please charge my Deposit Account No. 50-1407 in the amount of \$505.00 to cover the above fees. A duplicate copy of this sheet is enclosed. <input type="checkbox"/> The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. 50-1407 . A duplicate copy of this sheet is enclosed. NOTE: Where an appropriate time limit under 37CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status. SEND ALL CORRESPONDENCE TO:			
SOL SHEINBEIN G.E. EHRLICH (1995) LTD. C/O ANTONHY CASTORINA SUITE 207 2001 JEFFERSON DAVIS HIGHWAY ARLINGTON, VIRGINIA 22202, USA		<div style="text-align: center;">  SIGNATURE </div> <div style="text-align: center;"> SOL SHEINBEIN NAME </div> <div style="text-align: center;"> 25.457 REGISTRATION NUMBER </div> <div style="text-align: center;">  DATE </div>	

8. (Amended) The method according to claim 4, wherein said second genetic trait is of anti-shattering.

REMARKS

By this amendment, claims 6, 7, 8, 9, 10 and 12 have been amended to remove multiple dependency claims from the application as amended in PCT/IL00/00046 on January 24, 2000.

Attached herewith is a marked up version of the changes made to the specification and claims by the current amendment. The attached page is captioned "**VERSION WITH MARKS TO SHOW CHANGES MADE**".

Please charge any additional fees, if required, to Deposit Account 50-1407.

Respectfully submitted,

Sol Sheinbein
Attorney for Applicant
Registration No. 25,457

056637.07001
T09229" 2E25950

VERSION WITH MARKINGS TO SHOW CHANGES MADE

In the claims:

6. The method ~~or construct~~ according to ~~claims 4 or 5, respectively,~~
claim 4, wherein said second genetic trait is of abolished secondary dormancy.

7. The method ~~or construct~~ according to ~~claims 4 or 5, respectively,~~
claim 4, wherein said second genetic trait is of uniform or delayed ripening.

8. The method ~~or construct~~ according to ~~claims 4 or 5, respectively,~~
claim 4, wherein said second genetic trait is of anti-shattering.

9. The method ~~or construct~~ according to ~~claims 4 or 5, respectively,~~
claim 4 wherein said second genetic trait is of dwarfism.

10. The method ~~or construct~~ according to ~~claims 4 or 5, respectively,~~
claim 4 wherein said second genetic trait is selected from the group consisting
of seed stalk bolting, seed coat defects that facilitate uniform germination, root
storage promotion, biennial growth and non-flowering.

12. A genetically modified crop comprising the genetic construct of
~~claim 3 or 5.~~

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Int'l Appln No.: PCT/IL00/00046 §
Int'l Filing Date: 24 January 2000 §
Priority Date Claimed: 03 February 1999 §
Title: TRANSGENIC PLANTS §
Applicant: YEDA RESEARCH AND DEVELOPMENT §
CO. LTD. §
Attorney Docket: 00/20550 (Previously 34/52) §

RESPONSE

BOX PCT
Commissioner of Patents and Trademarks
Washington, D.C. 20231

Dear Sir:

This is a response to a Written opinion sent March 6, 2001, a response to which is due, and being made, within one month.

Claim 2 has now been canceled and the remaining claims renumbered. Enclosed herewith are replacement pages 35-37 replacing the corresponding pages of the application as filed.

PCT Article 33(3), Rejections

The Examiner has stated that claim 2 lacks an inventive step under PCT Article 33(3) as being obvious over Zemetra et al.

Claim 2 has now been canceled rendering moot the Examiners rejections with respect to this claim.

The Examiner also states that claims 3, 4 and 13 lack an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Williams et al.

The Examiner points out that Zemetra et al. teach a method of preventing introgression of desirable traits from a genetically engineered crop species into a closely related weedy species, but do not teach control elements which are not functional in weeds.

Williams et al. teach the use of genetic engineering for pollination control and the use of inducible promoters for controlled expression of desired genes.

The Examiner concludes that it would have been obvious to one of ordinary skill in the art to utilize the method of Zemetra et al. along with the inducible promoters taught by Williams et al. given the recognition by those of ordinary skill in the art that choice of means to prevent gene transmission or expression in weeds would have been the optimization of process parameters.

Applicant respectfully submits that this statement is in error as the cited combination of references does not provide one skilled in the art with the basis or motivation to practice the presently claimed invention.

The claimed invention relates to genetic mechanisms for mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop.

Claim 3 of the instant application relates to one possible mitigation method which is effected by introducing into a crop plant a control element inexpressible by the weed. Claims 4 and 13 relate to a construct utilizable by such method and a crop plant comprising such a construct, respectively.

As pointed out by the Examiner, Zemetra suggests mitigating introgression of a desirable trait by introducing the gene encoding such a desirable trait into a chromosome of a crop plant which would not introgress into a related weed.

In fact, Zemetra et al., suggests that the gene of choice be placed on a chromosome which, in theory, would not introgress into the weed; e.g., on the A or B genome of hexaploid wheat, and not on the D genome which is almost identical to the D genome of tetraploid *Aegilops cylindrica*.

In theory, such an approach would only work in two crop-weed pairs - wheat/*Aegilops* and oilseed rape (tetraploid AC) and *Brassica campestris* (C genome only).

Such mitigation of introgression approach would not be successful in crop-weed pairs which display close homology between respective genomes, such as, for example, rice-red rice and sunflowers-wild sunflowers.

In fact, numerous articles discuss the shortcomings of the method described by Zemetra et al. To this end, see, for example, Tomiuk J, Hauser TP, Bagger-Jørgensen R. 2000. A- or C-chromosomes, does it matter for the transfer of transgenes from *Brassica napus*. *Theoretical and Applied Genetics* 100: 750-4; Mikkelsen TR, Jensen J, Jørgensen RB. 1996. Inheritance of oilseed rape (*Brassica napus*) RAPD markers in a backcross progeny with *Brassica campestris*. *Theoretical and Applied Genetics* 92: 492-7 and Wang, ZN; Hang, A; Hansen, J; Burton, C; Mallory-Smith, CA; Zemetra, RS Visualization of A- and B-genome chromosomes in wheat (*Triticum aestivum* L.) x jointed goatgrass (*Aegilops cylindrica* Host) backcross progenies. *GENOME*, 43:1038-1044, 2000.

In sharp contrast, the claimed invention overcomes such a limitation by providing methods which do depend upon differences in genomic composition between crops and their respective weeds but rather takes advantage of variable gene expression and function. Thus, the methods of the claimed invention are not designed to prevent gene transfer but rather to prevent gene expression, function or alternatively to be detrimental to the weed when transferred.

09889737 072004
100220 4248850

In sharp contrast, the method described by Zemetra et al. attempts to block gene transfer, which as evident from the articles referenced above, is a near impossible task.

Williams et al. teach genetic engineering based methods of producing hybrid seeds in crop plants.

To produce such seeds, Williams et al. utilize construct systems for inducing male sterility which is reversible either through crossing or chemical induction of a counter-gene (inducible promoter recited on page 347, column 1, bottom paragraph).

Williams et al. do not teach nor do they suggest the use of such constructs or inducible promoters for mitigating introgression of a desired trait to a weed. In fact, it is possible that the chemically induced promoters described by Williams et al. are functional in weeds, since a crop restricted function of such inducible promoters is not described nor is it suggested by Williams et al.

Thus, since Zemetra et al., teaches chromosomal location of desired traits to prevent introgression and Williams et al. teach the use of inducible promoters for reversing male sterility, the cited references not only fail to provide teachings enabling or motivating one of skill in the art to practice the claimed invention, but also fail to provide any teachings that one skilled in the art would have a reasonable expectation of creating such methodology.

The Examiner further states that claim 1 lacks an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Kultnow et al. further in view of Williams et al.

The Examiner states that although Zemetra et al. do not teach the use of apomictic male-sterile plants, Kultnow et al. teach the use of apomictic plants for the propagation of plants without pollination, while Williams et al. teach the use of genetic engineering to create male sterile plants for pollination control.

Thus, the Examiner points out that it would have been obvious to combine the teachings referenced above given the benefits of pollination control taught by each reference.

Applicant respectfully submits that this statement is in error. As detailed below, the cited combination of references does not provide one skilled in the art with the basis or motivation to practice the presently claimed invention.

Zemetra et al. and Williams et al. were discussed above. Kultnow et al. describe the process of apomictic development and the use of molecular approaches that may lead to the isolation of apomictic genes which can be used for the generation of genetically identical seeds without fertilization. As is clearly stated by Kultnow et al. throughout the document, apomictic genes and plants can be used for agricultural production of improved hybrid crops by maintaining hybrid vigor while avoiding the limitations inherent to other prior art methods utilized. Kultnow et al. do not describe nor do they suggest the use of apomixis in mitigating introgression between crop plants and related weeds.

Thus, both Williams et al. and Kultnow et al. describe methods applicable for hybrid seed technology, and not for mitigating the effects of introgression.

Contrary to plants generated by the methods proposed by Williams et al. and/or Kultnow et al., the male sterile, apomictic plants generated according to the method of claim 1 of the present invention would not impregnate weeds.

It is the Applicant's strong opinion that the approach of the present invention which combines male sterility with apomictic crop plants is not rendered obvious by the combined teachings of Zemetra et al., Williams et al. and Kultnow et al., since such methodology was not suggested by the teachings thereof and since the advantages of such combination cannot be inferred therefrom by one of ordinary skill in the art.

09889737-072004

The Examiner also states that claims 5, 6, 9 and 13 lack an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Paterson et al. and further in view of Young.

Paterson et al. teach that seed shattering is a trait important to weeds, identify the chromosomal location of the gene and also caution against the introgression of desirable traits from cultivated crop species into weedy species.

Young teaches the isolation of an anti-shattering gene.

The Examiner points out that it would have been obvious to one of ordinary skill in the art to utilize the method of Zemetra et al. and to modify it by incorporating the anti-shattering gene taught by Young as suggested by Paterson et al.

Applicant respectfully submits that this statement is in error. As detailed below, the cited combination of references does not provide one skilled in the art with the basis or motivation to practice the presently claimed invention.

Claims 5, 6, 9 and 13 of the present invention relate to methods of mitigating the effects of introgression of a genetically engineered first genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, by co-engineering at least one copy of a genetically linked second genetic trait, encoding, for example, an anti-shattering function, in the crop which is innocuous or somewhat valuable to the crop yet deleterious to the weed.

In sharp contrast, Paterson et al. teach the use of a shattering gene for crop breeding. Although Paterson et al. "caution" against the use of transgenic sorghum, the use of shattering gene described thereby in an anti-shattering function (e.g., anti-sense) for the purpose of mitigating introgression is not described nor is it suggested.

Thus, It is the Applicant's strong opinion that an approach using an anti-shattering trait for mitigating introgression is not rendered obvious by the combined teachings of Zemetra et al., Paterson et al. and Young.

09889737 072001

The Examiner also states that claims 5-7 and 11-13 lack an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Paterson et al. and further in view of Snow et al.

Although Snow et al. analyses the ease with which crop to wild introgression can occur, and the fitness of the wild-crop hybrids and permeability of gene transfer. Snow et al. do not teach nor do they suggest methods useful for preventing such introgression.

As such, it is the Applicant's strong opinion that the combined teachings of Zemetra et al., Paterson et al. and Snow et al. do not render obvious claims 5-7 and 11-13 of the present invention.

The Examiner also states that claims 5, 6, 8 and 13 lack an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Paterson et al. and further in view of Klee et al. (U.S. Pat. No. 5,512,466).

The Examiner points out that Klee et al. teach crop plant transformation with a gene which delays fruit ripening, which when combined with the teachings of Zemetra et al. and Paterson et al. renders obvious claims 5, 6, 8 and 13 of the present invention.

Applicant would like to point out in this respect that since Klee et al. do not describe nor do they suggest the use of such ripening genes and plants expressing same for mitigating introgression, the combined teachings of Zemetra et al. in view of Paterson et al. and further in view of Klee et al. do not render obvious claims 5, 6, 8 and 13 of the present invention.

The Examiner also states that claim 10 lacks an inventive step under PCT Article 33(3) as being obvious over Zemetra et al. in view of Paterson et al. and further in view of Schaller et al.

Again, Applicant would like to point out that since Schaller et al. do not describe nor do they suggest the use of dwarfism inducing genes and plants expressing same for mitigating introgression, the combined teachings of Zemetra et al. in view of Paterson et al. and further in view of Schaller et al. do not render obvious claims 5, 6, 8 and 13 of the present invention.

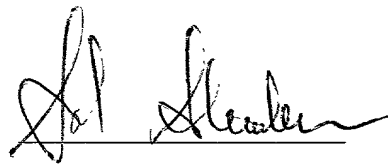
09889737.072001
T000270 4568850

Therefore, contrary to the Examiner's assertion, the prior art cited does not add much to the teachings of Zemetra et al. to suggest the presently claimed invention and as such it fails to provide one of ordinary skill in the art with motivation to practice the various aspects of the claimed invention.

In conclusion, it is the Applicant's strong opinion that the prior art method of mitigating introgression described by Zemetra et al. does not render obvious the novel methods of mitigating introgression described and claimed by the instant application simply because the prior art method is combined with teachings of components utilizable by the methods of the present invention, especially in cases in which teachings or suggestion for use of such components in mitigating introgression are clearly lacking.

In view of the above remarks it is respectfully submitted that claims 1-12 are non-obvious with respect to the prior art cited. Favorable examination is hence respectfully and earnestly solicited.

Respectfully submitted,

A handwritten signature in black ink, appearing to read 'Sol Sheinbein', written over a horizontal line.

Sol Sheinbein
Attorney for Applicant
Registration No. 25,457

Date: January 28, 2001.

09889737-072001
T000220" 4.5258859

WHAT IS CLAIMED IS:

1. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of producing apomictic seeds of said crop of a type which give rise to male sterile crop plants to thereby prevent introgression of the genetically engineered genetic trait of said crop to said weed and to reduce the weedy potential of the crop.

2. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of controlling the expression of the genetically engineered genetic trait in the crop by at least one control element which is inexpressible by the weed.

3. A genetic construct for genetically modifying a crop to express a genetically engineered genetic trait while mitigating the effects of introgression of the genetically engineered genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment encoding for said genetic trait and at least one additional nucleic acid segment including at least one control element which is expressible by the crop, yet which is inexpressible by the weed.

4. A method of mitigating the effects of introgression of a genetically engineered first genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of co-engineering at least one copy of a genetically linked second genetic trait in said crop, said second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

T00220" 2526850
09889737-072004

5. A genetic construct for genetically modifying a crop to express a genetically engineered first genetic trait while mitigating the effects of introgression of the genetically engineered first genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment encoding for said first genetic trait and at least one additional nucleic acid segment encoding a second genetic trait, said second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

6. The method or construct according to claims 4 or 5, respectively, wherein said second genetic trait is of abolished secondary dormancy.

7. The method or construct according to claims 4 or 5, respectively, wherein said second genetic trait is of uniform or delayed ripening.

8. The method or construct according to claims 4 or 5, respectively, wherein said second genetic trait is of anti-shattering.

9. The method or construct according to claims 4 or 5, respectively, wherein said second genetic trait is of dwarfism.

10. The method or construct according to claims 4 or 5, respectively, wherein said second genetic trait is selected from the group consisting of seed stalk bolting, seed coat defects that facilitate uniform germination, root storage promotion, biennial growth and non-flowering.

11. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed, the method comprising the step of cytogenetically selecting for or producing genetically engineered crop plants in which a gene or genes responsible for the genetic trait are

09889737-072001

genetically linked to an endogenous genetic trait of said crop, said endogenous genetic trait being deleterious to the weed.

12. A genetically modified crop comprising the genetic construct of claim 3 or 5.

0588977-076004
T00270" 268850

TRANSGENIC PLANTSFIELD AND BACKGROUND OF THE INVENTION

The present invention relates to a genetic mechanism for mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop and, more particularly, to a genetic mechanism for mitigating the effects of introgression of genetically engineered resistances of crops to weeds.

Crop domestication and weeds: During the prehistoric and historic processes of domestication of crops, farmers selected against a large number of traits that were valuable for wild species, but detrimental to agronomic practice. These differences between wild species and crops were further accentuated by selective breeding, and even more so by genetic engineering, which allowed introducing traits that were non-existent in the gene pool of the species, genus, family, or kingdom of the crop.

Concurrently with domestication; a few wild species evolved to fill the new ecological niches, the disturbed ecosystems known as farmers' fields (Baker, 1974; Holt, 1988; Turner, 1988). Only a few hundred of the tens of thousands of wild species have followed this evolutionary pathway from wild plant to widespread agricultural weed (Holm *et al.*, 1997). Thus, even though some weeds are closely related to crops or are even of the same species as the crops, they vary in a number of traits that distinguish them from wild species, as well as from the crop. These evolutionary processes are not static; indeed they are quite dynamic even on a human generation timescale (Baker, 1991). Changes in agricultural practices (drainage, fertilizer use, tillage and herbicide use) caused some pernicious weeds to return to being wild species, and some wild species to become weeds (Haas and Streibig, 1982). Crops can become "volunteer" weeds in the following crop, or even feral, and re-evolve some weedy traits. Some weeds have even introgressed new traits from conventionally-bred crops (wild barleys in barley have introgressed many new traits; wild sunflowers from sunflowers (Snow *et al.*, 1998). Worse, crops have introgressed weedy traits from related weeds e.g., poor oil quality in canola (Diepenbrock and Leon, 1988), and early bolting in sugar beets from weedy beets (Boudry *et al.*, 1998). These dynamic evolutionary events all occurred before the advent of transgenics.

Crops often possess conventionally-bred traits that would be advantageous to the weeds growing in their midst. Horizontal gene transfer

(introgression to totally unrelated species) occurs only rarely to other species within a genus, and even more rarely to species in closely related genera. Thus, even vital traits for weeds such as herbicide resistance have never passed horizontally from non transgenics (Torgersen, 1996), for example from wheat to grass weeds all in the family *Poaceae*. This lack of horizontal transfer allows the control of these related weeds in the crop. The weeds have had to evolve herbicide resistance from within their own genomes, and not by horizontal gene transfer.

The genetic distances between crop and weed were slightly enhanced with the advent of genetic engineering. Traits could be artificially forced horizontally into the crops to enhance cost-effectiveness of agriculture (higher yields, new products, resistances to insects, diseases, and to herbicides). Detractors of both the process of genetic engineering and its products have raised the possibilities that the engineered crops would become uncontrollable weeds, or that the genes would introgress into related weeds rendering them weedier, or into wild species, turning them into weeds (Kloppenburger, 1988; Goldberg *et al.*, 1990; Risler and Mellon, 1993). Hyper-generalizations were raised and terminology such as "superweeds" were coined (Kling, 1996). Calls were issued to prohibit or abandon all transgenic crops because of the possibilities of introgression of such traits into some weeds (Risler and Mellon, 1993). The fact that most crops have no interbreeding relatives in much of the world (Keeler *et al.*, 1996) did not allay fears for those crops. The issues made it to the popular press with kinky statements such as "The greatest danger of genetic engineering of plants may come from sex with weeds". The debate became as sterile as most of the interspecific hybrids generated using highly unnatural lab tricks to save the F₁ hybrids (Darmency, 1994).

Risk analysis and risk mitigation: Tomes were written on how to assess the risks of introgression – some with continuing generalizations and some discussing how and why this must be done case by case (Regal, 1994; Keeler *et al.*, 1996; Kareiva *et al.*, 1996; de Kathen, 1998; Williamson, 1993; Timmons *et al.*, 1996; Kjellsson *et al.*, 1998; Sindel, 1997; Gressel and Rotteveel, 1999, Galun and Breman, 1997; Krinsky and Wrubel, 1996). These discussion of the hazards and risk assessment do not discuss how biotechnologies can be used to mitigate the risk. No-one, including the governmental panels that permitted cultivating transgenic crops (Anonymous, 1994a, b, 1997) or those interested in regulatory aspects (Be *et al.*, 1996; Waters, 1996) has seemed to ask if there are ways to prevent

weeds from using any traits that may introgress from crops, in the few instances where one can be quite sure that introgression eventually will occur.

As further detailed hereinunder, a case by case analysis of where intra or interspecific introgressions between genetically engineered crops and weeds are possible shows that there are specific genetic possibilities for mitigation of interspecific introgression.

There is thus a widely recognized need for, and it would be highly advantageous to have, failsafe mechanisms to reduce the possibility of intra and interspecific introgressions between genetically engineered crops and weeds.

SUMMARY OF THE INVENTION

According to one aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of producing apomictic seeds of the crop of a type which give rise to male sterile crop plants to thereby prevent introgression of the genetically engineered genetic trait of the crop to the weed.

According to another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop having multiple genomes derived from different wild sources to a weed having a genome compatible with one of the multiple genomes, the method comprising the step of cytogenetically selecting for genetically engineered crop plants in which a gene or genes responsible for the genetic trait are localized on one or more of the multiple genomes of the crop which is not, or is far less, compatible with the genome of the weed.

According to still another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed, the method comprising the step of cytogenetically selecting for or producing genetically engineered crop plants in which a gene or genes responsible for the genetic trait are genetically linked to an endogenous genetic trait of the crop, the endogenous genetic being deleterious to the weed.

According to yet another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically

engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop , the method comprising the step of controlling the expression of the genetically engineered genetic trait in the crop by at least one control element which is inexpressible by the weed.

5 According to still another aspect of the present invention there is provided a genetic construct for genetically modifying a crop to express a genetically engineered genetic trait while mitigating the effects of introgression of the genetically engineered genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment
10 encoding for the genetic trait and at least one additional nucleic acid segment including at least one control element which is expressible by the crop, yet which is inexpressible by the weed.

According to an additional aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically
15 engineered first genetic trait of a crop to a weed and of mitigating a weedy potential of the crop , the method comprising the step of co-engineering at least one copy of a genetically linked second genetic trait in the crop, the second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

20 According to a further aspect of the present invention there is provided a genetic construct for genetically modifying a crop to express a genetically engineered first genetic trait while mitigating the effects of introgression of the genetically engineered first genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment
25 encoding for the first genetic trait and at least one additional nucleic acid segment encoding a second genetic trait, the second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

According to yet a further aspect of the present invention there is provided a crop genetically modified to include the above described genetic
30 constructs and to express the traits encoded thereby.

According to further features in preferred embodiments of the invention described below, the second genetic trait is of abolished secondary dormancy.

According to still further features in the described preferred
35 embodiments the second genetic trait is of uniform or delayed ripening.

According to still further features in the described preferred embodiments the second genetic trait is of anti-shattering of ripe seeds.

According to still further features in the described preferred embodiments the second genetic trait is of dwarfism.

According to still further features in the described preferred embodiments the second genetic trait is selected from the group consisting of seed stalk bolting, seed coat defects that facilitate uniform germination, root storage promotion, biennial growth and non-flowering.

The present invention successfully addresses the shortcomings of the presently known configurations by conceiving and providing a mechanism for mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop .

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is of a genetic mechanism which can be used for mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop. Specifically, the present invention can be used to prevent introgression of genetically engineered resistancies of crops to weeds.

The principles and operation of the present invention may be better understood with reference to the accompanying descriptions and examples.

Before explaining at least one embodiment of the invention in detail, it is to be understood that the invention is not limited in its application to the details of construction and the arrangement of the components set forth in the following description or illustrated in the drawings. The invention is capable of other embodiments or of being practiced or carried out in various ways. Also, it is to be understood that the phraseology and terminology employed herein is for the purpose of description and should not be regarded as limiting.

Generally, the nomenclature used herein and the laboratory procedures in recombinant DNA technology described below are those well known and commonly employed in the art. Standard techniques are used for cloning, DNA and RNA isolation, amplification and purification. Generally enzymatic reactions involving DNA ligase, DNA polymerase, restriction endonucleases and the like are performed according to the manufacturers' specifications. These techniques and various other techniques are generally performed according to Sambrook *et al.*, molecular Cloning--A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1989) which is incorporated herein by reference.

Other general references are provided throughout this document. The procedures and information therein are believed to be well known in the art and are provided for the convenience of the reader. All the information contained therein is incorporated herein by reference.

5 One of the greatest advantages of herbicide-resistant crops is that they allow control of closely-related weeds that have the same herbicide selectivity spectrum as the crop and could not be previously controlled. Similarly, an advantage of disease and insect resistant crops is that they can be grown where there are secondary hosts, often close relatives,
10 harboring the pests. Other resistant crops, e.g., cold resistance crops, are also of great advantage. Similarly, highly productive crops are advantageous, as are crops with modified product such as different types of starch and oils. Such and other genetic traits have been introduced into crops of various types by transgenics. These advantages of transgenics are
15 fine, if there is no introgression into a relative weed or if the crop itself does not become a "volunteer" weed in subsequent crops. Because the advantages of transgenics are so great in the above cases, both industry and the farmers are clamoring for the transgenics. They even do so when they know that introgression to weeds is imminent. However, if the gene for herbicide resistance introgresses, the farmers are just back to square one.
20 Considering the strong competition of red-rice with rice, and the magnitude of yield loss (Pantone and Baker, 1991), the desire of the farmers can be understood. One cannot state that this argument is wrong; just that there are a limited number of herbicide resistances one can engineer and thus a
25 limited number of times that one can return to square one.

Hence, while conceiving the present invention, the idea was realized to mitigate the risks of introgression of a genetically engineered trait of a crop to a weed by coupling the gene of choice having the desired trait in tandem constructs with "antiweediness" genes. This coupling can either be
30 physical, where the two genes are covalently linked prior to transformation or by the juxtaposition effect commonly achieved by co-transformation. Both will heretofore be termed "tandem", as the result in tightly linked genes. These would render weedy recipients or volunteer weeds less fit to act as competitors with crops, weeds and wild species. As further detailed
35 and exemplified hereinbelow, genes that prevent seed shatter, that prevent secondary dormancy, that dwarf the recipient and others would all be useful for that purpose, as they would often be beneficial or innocuous or somewhat valuable to the crop while detrimental to weeds, or to the crop

when it is a "volunteer" weed, i.e., when it becomes a weed in the following crop, or if it becomes feral.

As used herein in the specification and in the claims section that follows, the term "weed" includes undesirable plants growing wild, especially those growing on cultivated ground to the disadvantage of a crop, lawn, or flower bed. The term further includes various forms of the crop species that are undesirable to agriculture: feral forms that have escaped cultivation and have evolved weedy characters, other varieties of the crop that do not possess the same transgenes, and the transgenic crop when it is a volunteer weed in following crops.

Thus, according to one aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered trait of a crop to a weed and of mitigating a weedy potential of the crop. The method is effected by producing apomictic seeds of the crop of a type which give rise to male sterile crop plants, to thereby prevent introgression of the genetically engineered trait of the crop to the weed.

Male sterility in plants implies an inability to produce or to release functional (fertile) pollen. Male sterility in plants results in failure of formation or development of functional stamens, microspores or gametes.

From a structural/functional point of view, male sterility in plants may be divided into three categories which include (i) pollen sterility, wherein functional pollen grains are missing; (ii) structural (or staminal) male sterility, wherein male flowers or stamens are malformed and therefore non-functional, or missing altogether; and (iii) functional male sterility, wherein good and viable pollen is trapped in indehiscent anthers and thus prevented from functioning.

From a genetic point of view, male sterility in plants may also be divided into three categories which include (i) nuclear male sterility (NMS), also known in the art as genic or Mendelian male sterility, wherein male sterility is governed solely by one or more nuclear genes; (ii) cytoplasmic male sterility (CMS), wherein male sterility results due to a combined action of nuclear and cytoplasmic organelle (e.g., mitochondria or chloroplasts) genes; and (iii) non-genetic male sterility which is either chemically or mechanically (pollen removal) induced.

As further detailed hereinunder male sterility genes has been isolated and characterized. Such genes can be used to produce the apomictic seeds according to the present invention using genetic engineering techniques

09889737-072004

which are well known in the art, some of which are further described hereinunder.

According to another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered trait of a crop having multiple genomes derived from different wild sources to a weed having a genome compatible with one of the multiple genomes. The method is effected by cytogenetically selecting for genetically engineered crop plants in which a gene or genes responsible for the trait are localized on one or more of the multiple genomes of the crop which is not, or is far less compatible with the genome of the weed.

For example, the D genome of wheat is compatible with the D genome of *Aegilops cylindrica* (goatgrass) a problematic weed in the western plain states of the U.S. Transgenes can introgress from wheat to this *Aegilops* (Zemetra *et al.*, 1998). Likewise, transgenes easily introgress from the B genome of oilseed rape to many *Brassica* weeds and wild species (Darmency, 1994; Bing *et al.*, 1996; Brown and Brown, 1996; Jorgensen and Andersen, 1994; Kerlan *et al.*, 1993; Landbo *et al.*, 1996; Lefol *et al.*, 1996a, b Metz *et al.*, 1997; Mikkelsen *et al.*, 1997; Scheffler *et al.*, 1995). Selecting for wheat and oilseed rape transgenic plants in which no transgenes are integrated in the D or B genomes, will mitigate the possibility of introgression of the transgenic trait to *Aegilops cylindrica* and *Brassica* weeds and wild species, respectively.

Conventional methods of gene mapping in plants and the availability of genetic markers being specific to the chromosomes of the various genomes can be employed using well known and developed cytogenetic techniques to select for genetically engineered crop plants in which a gene or genes responsible for the trait are localized on one or more of the multiple genomes of the crop, which genome is not compatible with the genome of the weed.

According to yet another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically engineered trait of a crop to a weed. The method according to this aspect of the invention is effected by cytogenetically selecting for or producing genetically engineered crop plants in which a gene or genes responsible for the trait are genetically linked to an endogenous trait of the crop, the endogenous being deleterious to the weed.

According to still another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically

engineered trait of a crop to a weed and of mitigating a weedy potential of the crop . The method according to this aspect of the invention is effected by controlling the expression of the genetically engineered trait in the crop by at least one control element which is inexpressible by the weed.

5 Accordingly, the present invention provides a genetic construct for genetically modifying a crop to express a genetically engineered trait while mitigating the effects of introgression of the genetically engineered trait of the crop to a weed. The genetic construct includes a first nucleic acid segment encoding for the trait and at least one additional nucleic acid
10 segment including at least one control element which is expressible by the crop, yet which is inexpressible by the weed. These constructs can be tandemly produced, or the same effect can be achieved by co-transformation and co-lucis integration.

One example of a control element which is expressible by crop, yet is
15 inexpressible by weeds is the 35S promoter which was originally derived from cauliflower mosaic virus (CaMV) and which is silenced in plants infected by the virus (Al-Kaff *et al.*, 1998) - i.e., most cruciferous weed plants in the wild. One of the basic and fundamental mechanisms in the process of speciation (species formation) is loss or gain of genetic control
20 functions. It is therefor expected that a plurality of genetic control element will be functional in one species, yet non-functional in a closely related species.

According to still another aspect of the present invention there is provided a method of mitigating the effects of introgression of a genetically
25 engineered first genetic trait of a crop to a weed and of mitigating a weedy potential of the crop . The method according to this aspect of the present invention is effected by co-engineering at least one copy of a genetically linked second genetic trait in the crop, wherein the second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the
30 weed.

As used herein the term genetically linked refers to a genetic distance lower than 50 centiMorgan, preferably lower than 40 centiMorgan, more preferably lower than 30 centiMorgan, more preferably lower than 20 centiMorgan, more preferably lower than 10 centiMorgan, more preferably
35 lower than 5 centiMorgan, more preferably lower than 1 centiMorgan, most preferably in the range of 0 to 1 centiMorgan, wherein 0 centiMorgan refers to juxtaposed sequences.

T00270 269950

Accordingly, the present invention also provides a genetic construct for genetically modifying a crop to express a genetically engineered first genetic trait while mitigating the effects of introgression of the genetically engineered first genetic trait of the crop to a weed. The genetic construct includes a first nucleic acid segment encoding for the first genetic trait and at least one additional nucleic acid segment encoding a second genetic trait, the second genetic trait being innocuous or somewhat valuable to the crop, yet deleterious to the weed. The nucleic acid segment encoding the first genetic trait is preferably flanked on both sides by nucleic acid segments encoding the second genetic trait, to thereby reduce the risk of losing the second genetic trait due to mutation or crossing over.

However, it will be appreciated that in many cases while using conventional transformation techniques genetic traits carried on two different vectors end up integrating to the same locus.

Thus, according to a further aspect of the present invention there is provided a crop genetically modified to include the above described genetic constructs and to express the traits encoded thereby.

The second genetic trait is, as already pointed out, innocuous or somewhat valuable to the crop, yet deleterious to the weed. Numerous examples of such genetic traits are listed herein and are further discussed in the Examples section that follows.

One such trait is abolished secondary dormancy. Genetically abolished secondary dormancy would be neutral to the crop, or advantageous to some crops having some residual secondary dormancy, but deleterious to the weed. Tillage, crop rotation, and preplant use of herbicides, all standard practices, would control the uniformly-germinating weed seeds lacking secondary dormancy during the following season.

Another such trait is uniform or delayed ripening. For example, methods and constructs for controlling the ripening of fruits and vegetables are disclosed in U.S. Pat. No. 5,512,466 which teaches the expression of an ACC metabolizing enzyme in the fruit to inhibit the production of ethylene.

Yet another such trait is of anti-shattering of ripe seeds. Uniform ripening and anti-shattering genes would be a negative trait for weeds, neutral for uniformly ripening and non-shattering crops (e.g., rice), and positive, for example, for oilseed rape, which still has a shattering problem.

Still another such trait is dwarfism. Examples for dwarfism genes include genes relating to hormone production (Azpiroz *et al.*, 1998; Schaller *et al.*, 1998) as well as those dealing with shade avoidance, such as, but not

limited to, over expressed phytochrome genes which prevents recognition of shading and thus the plant remains short (Robson *et al.*, 1996). Additional examples are provided in the Examples section that follows.

Additional traits innocuous or somewhat valuable to the crop, yet deleterious to the weed include, but are not limited to, seed stalk bolting, seed coat defects that facilitate uniform germination, root storage promotion, biennial growth and non-flowering.

Thus, these anti-weediness traits are combined, according to the present invention, with the genetically engineered traits, which genetically engineered traits include, but are not limited to, traits imposing resistance to herbicides and plant pests and pathogens, traits imposing resistance to environmental conditions, such as, but not limited to, cold, salinity, etc., and traits affecting yield, ripening, as well as modified components such as starches and oils, etc. Examples of such traits for which genes has been isolated are summarized in many recent texts such as Galun and Breiman, 1997. Examples of such traits for which genes has yet not been isolated, however, their isolation is readily available, include shattering of seeds, precluding secondary dormancy and promoting biennial growth.

Once a gene responsible for a mitigating trait has been selected, it must be engineered for plant expression along with the trait which confer an advantage thereto. To introduce such genes into a plant, a suitable chimeric gene and transformation vector must be constructed. A typical chimeric gene for transformation into a plant will include a promoter region, a heterologous structural DNA coding sequences and a 3' non-translated polyadenylation site. A heterologous structural DNA coding sequence means a structural coding sequence that is not native to the plant being transformed. Heterologous with respect to the promoter means that the coding sequence does not exist in nature in the same gene with the promoter to which it is now attached. Chimeric means a novel non-naturally occurring gene which is comprised of parts of different genes. In preparing the transformation vector, the various DNA fragments may be manipulated as necessary to create the desired vector. This includes using linkers or adaptors as necessary to form suitable restriction sites or to eliminate unwanted restriction sites or other like manipulations which are known to those of ordinary skill in the art.

Promoters which are known or found to cause transcription of selected gene or genes in plant cells can be used to implement the present invention. Such promoters may be obtained from plants, plant pathogenic

bacteria or plant viruses and include, but are not necessarily limited to, the 35S and, 19S promoters of cauliflower mosaic virus (CaMV35S and CaMV19S), the full-length transcript promoter from the figwort mosaic virus (FMV35S) and promoters isolated from plant genes such as EPSP synthase, ssRUBISCO genes and promoters obtained from T-DNA genes of *Agrobacterium tumefaciens* such as nopaline and mannopine synthases. The particular promoter selected should be capable of causing sufficient expression to result in the production of an effective amount of the respective proteins to confer the traits.

Particularly useful promoters for use in some applications of the present invention are fruit specific promoters and the full-length transcript promoter from the figwort mosaic virus (FMV35S). The FMV35S promoter is particularly useful because of its ability to cause uniform and high levels of expression in plant tissues. The DNA sequence of a FMV35S promoter is presented in U.S. Pat. No. 5,512,466 and is identified as SEQ ID NO:17 therein. Examples of fruit specific promoters include the E8, E4, E17 and J49 promoters from tomato (Lincoln *et al.*, 1988), as well as the 2A11 promoter as described in U.S. Pat. No. 4,943,674.

The promoters used for expressing the genes according to the present invention may be further modified if desired to alter their expression characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene which represses the expression of ssRUBISCO in the absence of light, to create a promoter which is active in leaves but not in roots. The resulting chimeric promoter may be used as described herein. As used herein, the phrase "CaMV35S" or "FMV35S" promoter includes variations of these promoters, e.g., promoters derived by means of ligation with operator regions, random or controlled mutagenesis, addition or duplication of enhancer sequences, etc.

The 3' non-translated region contains a polyadenylation signal which functions in plants to cause the addition of polyadenylated nucleotides to the 3' end of an RNA sequence. Examples of suitable 3' regions are the 3' transcribed, non-translated regions containing the polyadenylation signal of the tumor-inducing (Ti) plasmid genes of *Agrobacterium*, such as the nopaline synthase (NOS) gene, and plant genes like the 7s soybean storage protein genes and the pea E9 small subunit of the RuBP carboxylase gene (ssRUBISCO).

The RNAs produced by a DNA construct of the present invention also preferably contains a 5' non-translated leader sequence. This sequence

can be derived from the promoters selected to express the genes, and can be specifically modified so as to increase translation of the mRNAs. The 5' non-translated regions can also be obtained from viral RNA's, from suitable eukaryotic genes, or from a synthetic gene sequence. The present invention is not limited to constructs wherein the non-translated region is derived from the 5' non-translated sequence that accompanies the promoter sequence. Rather, the non-translated leader sequences can be part of the 5' end of the non-translated region of the native coding sequence for the heterologous coding sequence, or part of the promoter sequence, or can be derived from an unrelated promoter or coding sequence as discussed above.

In a preferred embodiment according to the present invention, the vector that is used to introduce the encoded proteins into the host cells of the plant will comprise an appropriate selectable marker. In a more preferred embodiment according to the present invention the vector is a plant expression vector comprising both a selectable marker and an origin of replication. In another most preferred embodiment according to the present invention the vector will be a shuttle vector, which can propagate both in *E. coli* (wherein the construct comprises an appropriate selectable marker and origin of replication) and be compatible for propagation or integration in the genome of the plant organism of choice. In yet another embodiment, the construct comprising the promoter of choice, and the gene of interest is placed in a viral vector which is used to infect the cells. This virus may be integrated in the genome of the organism of choice or may remain non-integrated.

According to some embodiment of the present invention secretion of the protein or proteins out of the cell is preferred. In this embodiment the construct will comprise a signal sequence to effect secretion as is known in the art. For some applications, a signal sequence that is recognized in the active growth phase will be most preferred. As will be recognized by the skilled artisan, the appropriate signal sequence should be placed immediately downstream of the translational start site (ATG), and in frame with the coding sequence of the gene to be expressed.

Introduction of the construct into the cells is accomplished by any conventional method for transfection, infection or the like as is known in the art. In constructs comprising a selectable marker the cells may be selected for those bearing functional copies of the construct. If the plasmid comprising the gene of interest is episomal the appropriate selective conditions will be used during growth. Stable transfectants and stable cell

lines may be derived from the transfected cells in appropriate cases, in order to conveniently maintain the genotype of interest. Cell growth is accomplished in accordance with the cell type, using any standard growth conditions as may be suitable to support the growth of the specific cell line.

5 A DNA construct of the present invention can be inserted into the genome of a plant by any suitable method. Suitable plant transformation vectors include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, such as those disclosed by U.S. Pat. No. 4,940,838 and others. In addition to plant transformation vectors derived from the Ti or root-inducing (Ri) plasmids of *Agrobacterium*, alternative methods can be used
10 to insert the DNA construct of this invention into plant cells. Such methods may involve, for example, the use of liposomes, electroporation, chemicals that increase free DNA uptake, particle gun technology, and transformation using viruses. Methods for the introduction of vectors into maize, or other
15 monocot cells would include, but are not limited to, injection methods or microprojectile methods.

The construction of vectors capable of being inserted into a plant genome via *Agrobacterium tumefaciens* mediated delivery is known to those of ordinary skill in the art. Typical plant cloning vectors comprise
20 selectable and scoreable marker genes, T-DNA borders, cloning sites, appropriate bacterial genes to facilitate identification of transconjugates, broad host-range replication and mobilization functions and other elements as desired.

If *Agrobacterium* mediated delivery is chosen, once the vector has
25 been introduced into the disarmed *Agrobacterium* strain, the desired plant can then be transformed. Any known method of transformation that will work with the desired plant can be utilized.

Additional objects, advantages, and novel features of the present
30 invention will become apparent to one ordinarily skilled in the art upon examination of the following examples, which are not intended to be limiting. Additionally, each of the various embodiments and aspects of the present invention as delineated hereinabove and as claimed in the claims section below finds experimental support in the following examples.

EXAMPLES

Reference is now made to the following examples, which together with the above descriptions, illustrate the invention in a non limiting fashion.

Mitigation via failsafes

There are various failsafe mechanisms that can be used to mitigate the risk of introgression of traits between genetically engineered crops and weeds. Most of the discussion will describe the introduction of specific mitigating transgenes, but other mechanisms can also be envisaged, as described below.

Apomixis as a failsafe: One area that may develop is apomixis (Koltunow *et al.*, 1995), where seed is actually of vegetative origin. This is being developed to establish hybrid vigour without crosses. If apomictic varieties are pollen free, then genes cannot introgress into other species or into varieties of a crop. The lack of viable pollen is probably the only failsafe that would be acceptable to some detractors, who fear intervarietal movement of transgenes.

Cytogenetic Failsafes: Some crops such as wheat and oilseed rape are composed of multiple genomes derived from different wild sources (Kimber and Sears, 1997; U, 1935). In any given locale it is possible that only one of the genomes of the crop is identical to that of a related weed allowing easy gene transfer. For example: the D genome of wheat is compatible with the D genome of *Aegilops cylindrica* (bearded goatgrass) a problematic weed in the western plain states of the U.S. Transgenes can introgress from wheat to this *Aegilops* (Zemetra *et al.*, 1998). Likewise, transgenes easily introgress from the B genome of oilseed rape to many *Brassica* weeds and wild species (Darmency, 1994; Bing *et al.*, 1996; Brown and Brown, 1996; Jorgensen and Andersen, 1994; Kerlan *et al.*, 1993; Landbo *et al.*, 1996; Lefol *et al.*, 1996a, b Metz *et al.*, 1997; Mikkelsen *et al.*, 1997; Scheffler *et al.*, 1995). The further the genetic distances of between crop and weed in these crosses, the greater the needs for techniques such as embryo rescue to save hybrids abortion, and the greater the incidence of infertile offspring. The natural integration of a transgene on the D genome of wheat or the B genome of oilseed rape in interspecific crosses is quite simple. This should not be the case if the transgene is on the incompatible A or B genomes of wheat or the C genome

of oilseed rape. When the transgene is on one of the incompatible genomes, rare homologous recombination (crossing over) is required to integrate the transgene into stable and fertile offspring. Thus, cytogenetic mapping of transgenes, and releasing only those transgenic lines where the transgene is on genome incompatible with local weeds will lower the risk of introgression with weeds by orders of magnitude (Gressel and Rotteveel, 1999). Surprisingly, such risk lowering has not appeared among the requirements of regulatory authorities (Anonymous, 1994a, b, 1997; Be *et al.*, 1996; Waters, 1996).

Transgenetic Mitigation (TM)

The concept of using genetic engineering to mitigate any positive effects transgenes may confer is based on three premises:

i. Tandem constructs of genes act genetically as tightly-linked genes and their segregation from each other is exceedingly rare.

ii. There are traits that are either neutral or positive for a crop that would be deleterious to a typical or volunteer weed, or to a wild species.

iii. Because weeds have a strong competition amongst themselves, and have a large seed output of weeds, even mildly deleterious traits are quickly eliminated from populations.

Thus, if the gene of choice being engineered into a crop is flanked on either side by a TM gene in a tandem construct, the overall effect would be deleterious to weeds introgressing the construct from a crop. Even if one of the TM genes mutates, is deleted, or crosses over, the other flanking TM gene will remain, providing mitigation.

One TM trait has already been put into transgenic crops, albeit inadvertently: the use of the 35S promoter in gene constructs of desirable traits in oilseed rape (*Brassica napus*). The 35S promoter was originally derived from cauliflower mosaic virus (CaMV). This promoter is silenced in plants infected by the virus (Al-Kaff *et al.*, 1998) - i.e., most cruciferous weed plants in the wild. Thus, a wild and weedy related *Brassica* that introgresses herbicide resistance from oilseed rape is suddenly herbicide sensitive in a CaMV-infected weed. One could consider the advertent use of such promoters.

Other TM traits that could be used are best visualized when observing the differences between crops and weeds. This is best illustrated with two cases: (i) rice and weedy red rice (both *Oryza sativa*) as well as

rice with wild rices *Oryza* spp.; (ii) oilseed rape (*Brassica napus*) and feral and weedy Polish rape/wild radish *B. campestris* = *B. rapa*, as summarized below.

Seed dormancy: Weed seeds typically have secondary dormancy with seeds from one harvest germinating bit by bit throughout the following season, and over a number of years (Vleeshouwers *et al.*, 1995). This evolutionary trait is considered to be a risk-spreading strategy that maximizes fitness while reducing losses due to sib competition (Hyatt and Evans, 1998; Lundberg *et al.*, 1996). Staggered secondary dormancy prevents all the weeds from being controlled by tillage before the crop is planted, or controlled by tillage or herbicides during crop rotation. Rare mutants lacking secondary dormancy were selectively propagated during crop domestication, as the loss of secondary dormancy is desirable to the farmer, who wants uniform germination after planting the crop. Crop seed that germinates uniformly after planting gives a uniform harvest. This can well be seen when comparing crops with their weedy progenitors and relatives (Ling-Hwa, 1997). Genetically abolishing secondary dormancy would be neutral to the crop, but deleterious to the weed. Tillage, crop rotation, and preplant use of herbicides, all standard practices would control the uniformly-germinating weed seeds lacking secondary dormancy during the following season.

Ripening and shattering: Weeds disperse their seed over a period of time and much of the ripe seed “shatters” to the ground. This ensures replenishment of the soil seed bank. A proportion of the weed seed is harvested with crop seed, contaminating crop seed, facilitating weed dispersal to wherever the crop seed will be grown. Weeds have evolved morphological and phenological “mimicries” to the crop seed (Barrett, 1983; Gould, 1991) necessitating continual evolution and refinement of seed cleaning techniques to remove the contaminating weed seed. Crop varieties have been selected for non-shattering, but recently domesticated crops such as oilseed rape still suffer from shattering (Simon, 1994; Prakash, 1988; Price *et al.*, 1996). The first problem in domestication is control of shattering (Young, 1991; Levy, 1985). In addition to the loss of yield, the shattering of crop seed causes the crop to be a volunteer weed in the following crop (Lutman, 1993).

Uniform ripening and anti-shattering genes would be a negative trait for the weeds, neutral for rice (because it ripens uniformly and does not shatter easily after thousands of years of selection), and positive for oilseed

rape, which still has a shattering problem. The addition of anti-shattering genes in a TM construct could also prevent cultivated oil seed rape from becoming a volunteer weed problem as well as being a TM gene. Crop seed contaminated with low levels of weed seed are typically used for feeding or processing, only weed free "certified" seed is sown.

Dwarfing:

Dwarfing has been especially valuable in generating "green revolution" crops, but also has value in normal cropping situations. The green revolution in rice and wheat is based on a modification of the harvest index, the ratio of grain to straw. For millenia these crops had been selected for height, to outgrow weeds, limiting the photosynthate available for grain. Weed evolution continued apace, giving rise to taller weeds. The advent of selective herbicides to kill weeds allowed for genetic dwarfing of these crops resulting in more seed harvest and less straw. Some of these dwarfing genes are tightly linked to genes reducing general yield potential. Still, the lowering of height, precluding the concomitant problem of tall plants "lodging" (falling over), and increased yield, especially after fertilizer use (which previously promoted lodging), allowed countries like India to become self sufficient, despite population increase.

Various new systems of genetically engineered height reduction are being introduced. These include genes relating to hormone production (Azpiroz *et al.*, 1998; Schaller *et al.*, 1998) as well as those dealing with shade avoidance. Much of stem elongation is in response to shading. This is advantageous when competing with other species, but not in a weed-free crop stand where only siblings are competing. The overexpression of specific phytochrome genes prevents recognition of shading and thus the plant remains short (Robson *et al.*, 1996). This is advantageous for a crop and could also be used where the present dwarfing genes prevent obtaining the highest yields. This trait would be disadvantageous for a weed that must compete with the crops; it would be shaded over by the crop.

QTLs: QTLs or other unidentified genes that have been shown to provide general weediness characters are known (Paterson *et al.*, 1995). QTLs have also been identified controlling dormancy (Van der Schaar *et al.*, 1997) and for the late stages of gibberellin biosyntheses (Lange *et al.*, 1997), which might relate to stem dwarfing or seed stalk bolting.

Other TM traits: One could envision other traits that would be disadvantageous to weeds but neutral to crops. These include removing seed coat characters that allow weed seeds to pass through animal digestive

tracks intact and then be dispersed. Genes that promote root storage would be advantageous to cultivated beets but detrimental to annual wild beets. Genes that prevent bolting (i.e., promote biennial growth) would be excellent for carrots, celery, cabbage, lettuce, bee, and related crops, but would be highly deleterious to related weeds. Anti-flowering genes would prevent introgression of genes from potatoes into wild Andean relatives (the only place where hazards of introgression from potatoes exist to weeds). They would also prevent volunteer potatoes arising from true seed, where this is a problem. Sorghum provides an interesting case. Crosses between cultivated sorghum (*S. bicolor*) and johnsongrass (*S. halapense*) gave rise to sterile, perennial, vegetatively-propagating plants (Baker, 1991). The hybrids have thick rhizomes storing large amounts of material. Various of the QTL's reported by Peterson *et al* (1995) could severely decrease the fitness of any hybrids that form.

Presumably, comparisons of crop traits with weed traits will lead to finding other TM traits.

Balancing desirous transgenic traits with TM traits

There is considerable debate about the advantages that would accrue to weeds from the primary transgenic traits. Resistance by modified site of the herbicide binding site to its target should only confer an advantage when the herbicide is used. It is unknown whether there would be pleiotropic advantages of herbicide resistance due to introducing genes for metabolic inactivation of herbicides. Insect, or pathogen resistances would clearly provide an advantage to a weed, if the weed does not already have these resistances, and is affected by the pest. Many pest resistances were bred out of wild species during domestication; the chemicals weeds use to alleviate pest problems often taste bad or are toxic to mammals.

Still, let us assume that the primary transgenic trait confers an advantage to a weed; how much will TM traits actually mitigate that advantage? Weeds are not only highly competitive with crops, they are competitive with weeds of other species as well as within their own species. Seed-producing weeds often produce thousands of seeds, in steady state conditions, to replace a single plant, suggesting extreme competition to be the replacement; the selection for high competitive fitness is intense. This has dual implications in the discussed situation. A weed that introgresses any transgenic trait, will proliferate and spread through a population very quickly, even if it has fitness advantage that is marginally positive (Thill

and Mallory-Smith, 1997; Crawford *et al.*, 1997). Conversely, one can balance the disadvantage of TM traits against the advantage of the primary trait. This must be done in both in the presence and absence of the reason for using the primary trait. The primary trait only provides an advantage when it is needed; when there is pressure from the pest, herbicide, or stress. Some primary traits that were not meant to confer any plant protective advantage to the crop might still do so when introgressed into a weed, e.g., changed oil or starch composition. Membrane lipid compositions do change in response to temperature and other stresses, and the ability to adapt to certain environments might be enhanced or decreased (Cooper and Raybould, 1997; Linder, 1998); a plant with a modified starch might be less palatable to insects or less digestible by fungi, and thus have an advantage. In the absence of the factor making the primary trait desirable it should not have any advantage, accentuating the utility of the TM traits. Indeed, when the primary selector is not present, the primary transgenic trait can be disadvantageous; this has been demonstrated with one herbicide resistance gene (Bergelson *et al.*, 1996). Many herbicides have a short residual effect and when they are not present, there is no advantage to having transgenic herbicide resistance. Similarly, while some insect pests disease pathogens are continually present, there are many others that appear in damaging levels only in certain seasons, or even only once every few years, when the climatic conditions are just right for their causing pandemics. If resistance to a disease or insect has only occasional value, and the resistance mechanism has a fitness penalty when not needed, the value of the primary gene is depleted, and TM genes will elicit a net decrease in fitness.

Likelihood of TM genes segregating from advantageous genes

Often, the expression of one gene of a tandem construct is lost in the primary transgenic plant, and sometimes the expression is lost after a few generations. The reasons for these losses are not always clear nor relevant for this discussion, as only stabilized progeny of transformants are released to agriculture. If all traits of a tandem construct are expressed after 4-5 generations of development, it is fair to consider it stable, i.e., as stable as any native, tightly-linked adjacent genes. The likelihood of the TM and desired trait segregating from each other is infinitesimally lower than for the TM trait being inactivated by a mutation. Thus, if the mutation frequency of inactivation of the TM gene is 10^{-6} to 10^{-7} , that would be the frequency of the loss of the TM trait as the frequency of crossing over is many orders

of magnitude lower. If the frequency of crossing over of tightly linked traits is an unacceptably high risk, then one can consider using two TM genes, flanking the primary gene on either side. The frequency of loss of two genes TM can be predicted as 10^{-12} to 10^{-14} . Each TM trait should work in a balance with the primary trait, and where the primary gene gives a strong advantage to a weed, it might be necessary to have more than one TM trait in a construct. The risk of losing a TM trait can be further decreased by combination with a cytogenetic failsafe, where it is available. If the tandem construct is located on a non-homologous chromosome (where such exist), then only rare homologous recombination can move it. As there is no selective advantage to losing the TM trait on the non-homologous chromosome, one can compound the frequency of likelihood of homologous recombination with the frequency of loss of the TM trait(s).

Do TM genes have to give 100% safety: No risk situations are impossible but how low a risk do we need to attain? This is really a question for regulators, but when they do deliberate the issue they must ask; when will the related weed evolve the trait in question by natural means. This is best illustrated by an example: one company in the United states engineered resistance of ALS (acetolactate synthase)-inhibiting herbicides into sunflowers (*Helianthus annuus*) but never even field tested them, because of fear that the gene might introgress into weedy wild sunflowers (also *H. annuus*), prevalent in the mid western plain states. Natural introgressions between this crop and related weed are common and cause problems (Arias and Riesenbergs, 1994; Whitton *et al.*, 1995). The mutations conferring resistance to ALS-inhibiting herbicides are naturally prevalent in plant populations in a frequency of one in a million, and ALS-inhibiting herbicides are widely used. Wild sunflowers have recently evolved resistance to ALS-inhibiting herbicides in monoculture cropping (White *et al.*, 1998). If a TM construct had been inserted in tandem with an ALS gene, with a likelihood of segregating of 10^{-10} , then the likelihood of getting ALS-resistant wild sunflowers would not have appreciably been changed by introgression from transgenics. Wild sunflowers could have been controlled in sunflower fields, lessening the possibilities of natural introgressions. Such analyses should be made wherever possible.

TM traits which are available as gene sequences

Some of the traits suggested for use as TM genes are just known to exist as named traits that are inherited, others are also mapped as genes to

positions on various chromosomes, and a few are actually characterized as sequenced genes. Thus, not all TM traits are immediately available for insertion in tandem constructs. Still, there can be many different ways for a plant to confer a TM trait, and thus, more than one gene might be available.

5 Many of the TM traits be introgressed into the crops are not yet available as genes for transformation, suggesting that efforts be instituted to isolate TM genes. Gene hunting to a large extent is no longer driven by a desire for basic knowledge; it is heavily driven by a perceived utility of the gene. Now that the TM genes have been given a value, they might be
10 rapidly sought and found, especially with the availability of transposon tagging as a method for hunting and fishing.

An interim solution may be to select transformants that have randomly introgressed the primary gene to a point in close linkage to a TM trait. Many of these TM traits are already mapped in major crops, such as
15 dormancy in rice (Wan *et al.*, 1998; Lin *et al.*, 1998). The closer the linkage distance, the lesser the likelihood of segregation. In the future, when technologies are available for site specific insertion of genes into chromosomes, the primary genes could be spliced close to TM genes, without having to know the sequence of the TM gene.

20 **Secondary dormancy:** While the genetics of secondary dormancy has been described, it is not clear which genes actually control it (Li and Foley, 1997; Khan, 1998). In important weeds such as *Avena fatum*, the lack of dormancy is dominant (Foley and Fennimore, 1998) such that this, as a transgene on the domesticated oats (*A. sativa*) would turn wild oats into
25 a less fit weed. Unfortunately, *Arabidopsis*, the typical source for genes, has already been sufficiently domesticated that it is unlike cruciferous weeds; the lab strains no longer have strong secondary dormancy (Van der Schaar *et al.*, 1997). An *Arabidopsis* mutant that is insensitive to abscisic acid, lacks secondary dormancy (Steber *et al.*, 1998). Perhaps a way to find
30 more genes is to use the genetic differences between wild *Arabidopsis* strains and the lab strains presently used, as is being done in other instances (Auckerman *et al.*, 1997).

An endo- β -mannanase has been shown to be part of wall softening during seed germination (Bewley, 1997). Such a gene could overcome
35 secondary dormancy in weeds, while accelerating the rate of primary germination in crops, itself a valuable trait.

Much has been published on the physiology of secondary dormancy and how its causes vary among species. In some cases it is due to

impervious seed coats and in others due to various inhibitors found in the seed.

Shattering: Physiologically, one way to avoid seed shattering is to have uniform ripening. Early maturing soybean and oilseed rape seeds on indeterminate continuously flowering varieties typically shatter. Thus, determinacy, with its single uniform flush of flowering is one method to prevent shattering, but this often shortens the season, reducing yield. The hormonology of the abscission zone controls whether shattering will occur and it is possible that if cytokinins are overproduced, then shattering will be delayed. In sorghum the genes that control shattering seem to be on QTLs (Paterson *et al.*, 1995) as discussed above.

Stature limitation:

Vertical deprivation (dwarfing) has proven itself as a desirable trait in crops due to the increase in harvest index by virtue of having less stem and more seed. The genes used so far seem to have an unknown function. Still many genes are known, via their physiological role, that could control height.

Gibberellins: It is well known that preventing the biosyntheses of gibberellins reduces the height. The genetic relationships between some dwarfing genes and gibberellin biosyntheses has been elucidated (Webb *et al.*, 1998). This is the basis of many chemical dwarfing agents used commercially to lower stature and prevent lodging of wheat. The enzymes and genes controlling various steps in gibberellin biosyntheses are also known. Copalyl diphosphate synthase, ent-kaurene synthase, and ent-kaurene oxidase are responsible for early stages in the biosynthesis of all gibberellins (Smith *et al.*, 1998; Yamaguchi *et al.*, 1998, Hedden and Kamiya, 1997; Lange, 1998; Helliwell *et al.*, 1998). *Arabidopsis* mutations bearing mutations in any of them are dwarfed, with the dwarfing being reversible by gibberellin treatment. Overexpression of a gene coding for ent-kaurene synthase, causing co-suppression mimicked the mutant phenotype.

Some processes, such as flower stalk bolting, are controlled by specific gibberellins; in radish, GA₁ and GA₄ are responsible for flower stalk bolting (Nishijima *et al.*, 1998). It may be necessary to characterize the genes coding for the monooxygenases and dioxygenases that are responsible for these later steps (Hedden, 1997). Some of these genes have been isolated as well (Kusaba *et al.*, 1998).

Brassinosteroids: This new group of hormones also causes elongation of stems in many plant species, and their absence results in dwarf plants. A 22 α -hydroxylase cytochrome P450 has recently been isolated that controls a series of these steps in brassinosteroid biosynthesis (Choe *et al.*, 1998), and plants missing the enzyme are dwarfed (Azpiroz *et al.*, 1998). Additionally, suppressive overexpression of a sterol C24-methyl transferase also causes dwarfing (Schaller *et al.*, 1998).

Shade avoidance:

Various forms of the pigment phytochrome interact to detect whether a plant is being shaded (Smith and Whitelam, 1997; Devlin *et al.*, 1998; Torii *et al.*, 1998; Auckerman *et al.*, 1997). Phytochrome recognition of shading leads to stem elongation, which is unneeded in a weed-free crop. The engineering of suppressive overexpression constructs of one of these phytochromes led to plants that did not elongate as a results of shading (Robson *et al.*, 1996). Much of the gene isolation has been from *Arabidopsis*, yet the suppressive overexpression was active in dwarfing tobacco.

TMs for vegetatively propagated crops: The genes for pollen sterility (Williams, 1995) would clearly be the simplest way to render potato to be without true seeds. Such potatoes could not become volunteer weeds (from true seeds), or have pollen that introgresses into other potato varieties or into wild related Andean species (Love, 1994; Eijlander and Steikema, 1994).

TMs for biennial crops: Biennial crops such as beets, carrots, etc. usually require a period of cold vernalization before flower stalk formation (bolting). At the end of the vernalization there is typically a burst of endogenous gibberellin biosynthesis, which induces stalk elongation, and exogenous gibberellins can often replace the cold requirement. Possibly, bolting could be suppressed by including a TM antisense or suppressive overexpression construct for one of the enzymes of gibberellin biosynthesis, both on crop and of related weed. These genes are known (see above).

Although the invention has been described in conjunction with specific embodiments thereof, it is evident that many alternatives, modifications and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all such alternatives, modifications and variations that fall within the spirit and broad scope of the appended claims. All publications cited herein are incorporated by reference in their

References:

1. Al-Kaff, N.S., S. N. Covey, M.M. Kreike, A.M. Page, R. Pinder and P.J. Dale. , 1998. Transcriptional and postranslational plant gene silencing in response to a pathogen. *Science* 279:2113-2115.
2. Anon., 1994a. Assessment criteria for determining environmental safety of plants with novel traits. Directive Dir. 94-08 Plant Products Division, Agriculture and Agri-Food Canada, Nepean Ontario. (<http://www.cfia-acia.agr.ca/English/food/pbo/dir9408.html>).
3. Anon., 1994b. The biology of *Brassica napus* L (canola/rapessed). Directive Dir. 94-09 Plant Products Division, Agriculture and Agri-Food Canada, Nepean Ontario. (<http://www.cfia-acia.agr.ca/English/food/pbo/dir9409.html>).
4. Anon., 1997 Consensus document on the biology of *Brassica napus* L. (oilseed rape). Series on the harmonization of regulatory oversight in biotechnology No. 7. Environ. Directorate. Org. for Econ. Co-op. and Devel., Paris.
5. Arias, D.M. and L.H. Riesenbergs. , 1994. Gene flow between cultivated and wild sunflowers. *Theor. Appl. Genet.* 89:655-660.
6. Aukerman, M.J., M. Hirschfeld, L. Wester, M. Weaver, T. Clack, R.M. Amasino and R.A. Sharrock. , 1997. A deletion in the PHYD gene of the *Arabidopsis* Wassilewskija ecotype defines a role for phytochrome D in red/far-red light sensing. *Plant Cell* 9:1317-1326.
7. Azpiroz, R., Y. Wu, J. C. LoCascio and K.A. Feldmann., 1998. An *Arabidopsis* brassinosteroid-dependent mutant is blocked in cell elongation. *Plant Cell* 10: 219-230.
8. Baker, H.G., 1974. The evolution of weeds. *Ann. Rev. Ecol. Sys.* 5:1-24.
9. Baker, H.G., 1991. The continuing evolution of weeds. *Econ. Bot.* 45:445-449.
10. Barrett, S.C.H., 1983. Crop mimicry in weeds. *Econ. Bot.* 37:255-282.
11. Be, D.B., S.G. Rogers, T.B. Stone, and F.S. Serdy. , 1996. Herbicide tolerant plants developed through biotechnology: Regulatory considerations in the United States. p. 341-346. In: S.O. Duke (ed.), *Herbicide resistant crops: Agricultural, environmental, economic, regulatory, and technical aspects*. CRC Press, Boca Raton.

12. Bergelson, J., C.B. Purrington, C.J. Palm, and J.C. Lopez-Guitierrez. , 1996. Costs of resistance – a test using transgenic *Arabidopsis thaliana*. Proc. Roy. Soc. Lond. B. Biol. Sci. 262: 1659-1663.
13. Bewley, J. D. , 1997. Breaking down the walls - a role for endo-beta-mannanase in release from seed dormancy? Trends Plant Sci. 2: 464-469.
14. Bing, D.B., R.K. Downey, and G.F.W. Rakow. , 1996 Assessment of transgene escape from *Brassica rapa* (*B. campestris*) into *B. nigra* or *Sinapis arvensis*. Plant Breed. 115:1-4.
15. Boudry, P., M. Mirchen, P. Saumitou-Laprade, H. Vernet, and H. Van Dijk., 1993. The origin and evolution of weed beets: consequences for the breeding and release of herbicide-resistant transgenic sugar beets. Theor. Appl. Genet. 87:471-478.
16. Brown, J., and A.P. Brown., 1996. Gene transfer between canola (*Brassica napus* L. and *B. campestris* L.) and related weed species. Ann. Appl. Biol. 129:513-522.
17. Choe, S. B.P. Dilkes, S. Fugioka, S. Takatsuto, A. Sukarai, and K.A. Felmann. , 1998. The DWF4 gene of *Arabidopsis* encodes a cytochrome P450 that mediates multiple 22 α -hydroxylation steps in brassinosteroid biosynthesis. Plant Cell 10:231-144.
18. Conner, A.J., and P.J. Dale. , 1996. Reconsideration of pollen dispersal data from field trials of transgenic potatoes. Theor. Appl. Genet. 92:505-508.
19. Cooper, J.I. and A.F. Raybould. , 1997. Transgenes for stress tolerance: consequences for weed evolution. Brighton Crop Protect. Conf. - Weeds. pp. 265-272.
20. Crawford, J., G. Squire, and D. Burn. , 1997. Modeling spread of herbicide resistant oilseed rape. In: A.J. Gray, C. Glidden and F. Amjee (eds.). Environmental Impact of Genetically Modified Crops. Dept. of Environment, London, (in press).
21. Crawley, M.J., R.S. Hails, M. Rees, D. Kohn, and J. Buxton. , 1993. Ecology of transgenic oilseed rape in natural habitats. Nature 363:620-623.
22. Darmency, H., 1994. The impact of hybrids between genetically modified crop plants and their related species: introgression and weediness. Mol. Ecol. 3:37-40.

23. DeKathen, A., 1998. The debate on risks from plant biotechnology: the end of reductionism? *Plant Tissue Culture and Biotechnology* 4: 136-148.
24. Devlin, P.F., S.R. Patel and G.C. Whitlam., 1998. Phytochrome E influences internode elongation and flowering time in *Arabidopsis*. *Plant Cell* 10:1479-1488.
25. Diepenbrock, W. and J. Leon. , 1988. Quantitative effects of volunteer plants on glucosinolate content in double-low rapeseed (*Brassica napus* L.): a theoretical approach. *Agronomie* 8:373-377.
26. Eijlander, R., and W.J. Stiekema., 1994. Biological containment of potato (*Solanum tuberosum*): outcrossing to the related wild species black nightshade (*Solanum nigrum*) and bittersweet (*Solanum dulcamara*). *Sex Plant Reprod.* 7: 29-40.
27. Foley, M. E., and S.A. Fennemore. , 1998. Genetic basis for seed dormancy. *Seed Sci. Res.* 8:173-182.
28. Galun, E. and A. Breiman. , 1997. *Transgenic Plants*. Imperial College Press, London, 376 pp.
29. Goldberg, R., J. Rissler, H. Shand, and C. Hassebrook. , 1990. *Biotechnology's bitter harvest*. Environmental Defense Fund, New-York.
30. Gould, F. , 1991. The evolutionary potential of crop pests. *Amer. Sci.* 79: 496-507.
31. Gressel, J., 1997. Genetic engineering can either exacerbate or alleviate herbicide resistance. *Proc. 50th New Zealand Plant Protection Conf.* p. 298-306.
32. Gressel, J., and A.W. Rotteveel. , 1999. Risks from Biotechnologically-Derived Herbicide-Resistant Crops: Decision Trees for Assessment. *Plant Breeding Rev.* (in press).
33. Haas, H., and J.C. Streibig. , 1982. Changes in weed distribution patterns as a result of herbicide use and other agronomic factors. p. 57-80. In: H.M. LeBaron and J. Gressel (eds.), *Herbicide resistance in plants*. Wiley, New-York.
34. Hedden, P. , 1997. The oxidases of gibberellin biosynthesis: Their function and mechanism. *Physiol. Plant.* 101:709-719.
35. Hedden, P., and Y. Kamiya. , 1997. Gibberellin biosynthesis: Enzymes, genes and their regulation. *Ann. Rev. Plant Physiol. Plant Mol. Biol.* 48: 431-460.

36. Helliwell, C.A., C.C. Sheldon, M.R. Olive, A.R. Walker, J.A.D. Zeevaart, W.J. Peacock and E.S. Dennis., 1998. Cloning of the *Arabidopsis* ent-kaurene oxidase gene GA3. Proc. Natl. Acad. Sci. U.S.A. 95:9019-9024.
37. Holm, L., J. Doll, E. Holm, J. Pancho, and J. Herberger. , 1997. Worlds weeds: Natural histories and distributions. Wiley, New York.
38. Holt, J.S. , 1988. Ecological and physiological characteristics of weeds. P: 7-23. In: M.A. Altieri and M. Liebman (eds.), Weed Management in Agroecosystems: Ecological Approaches. CRC Press, Inc. Boca Raton, Florida.
39. Hyatt, L. A., and A.S. Evans. , 1998. Is decreased germination fraction associated with risk of sibling competition? OIKOS. 83: 29-35
40. Jorgensen, R.B., and B. Andersen. , 1994. Spontaneous hybridization between oilseed rape (*Brassica napus*) and weedy *B. campestris*: a risk of growing genmodified oilseed rape. Am. J. Bot. 81: 1620-1626.
41. Kareiva, P., I.M. Parker, and M. Pascual. , 1996. Can we use experiments and models in predicting the invasiveness of genetically engineered organisms? Ecology 77:1670-1675.
42. Keeler, K.H., C.E. Turner, and M.R. Bollick. , 1996. Movement of crop transgenes into wild plants. p. 303-330 In: S.O. Duke (ed.), Herbicide resistant crops: Agricultural, environmental, economic, regulatory, and technical aspects. CRC Press, Boca Raton.
43. Kerlan, M.C., A.M. Chevre, and F. Eber, F., 1993. Interspecific hybrids between a transgenic rapeseed (*Brassica napus*) and related species: cytogenetical characterization and detection of the transgene. Genome 36:1099-1106.
44. Khan, A. A. , 1997. Quantification of seed dormancy: Physiological and molecular considerations. Hortsci. 32: 609-614.
45. Kimber, G., and E.R. Sears. , 1987. Evolution of the genus *Triticum* and the origin of cultivated wheat. In: E.G. Heyne (ed.), Wheat and wheat improvement. Agronomy Monograph No. 13. ASA-SCCA-SSSA, Madison WI, pages. 154-164.
46. Kjellsson, G., V. Simonsen, and K. Ammann, (eds.), 1994. Methods for risk assessment of transgenic plants. Vol. 2. Pollination, gene transfer and population impacts. Birkhaeuser, Basel.
47. Kling, J., 1996. Could transgenic supercrops one day breed superweeds? Science 274:180-181.

48. Kloppenburg, J., Jr. , 1988. First the seeds: The political economy of plant biotechnology, Cambridge Univ. Press, Cambridge.
49. Koltunow, A.M., R.A. Bicknell, and A.M. Chaudhury., 1995. Apomixis: Molecular strategies for the generation of genetically-identical seeds without fertilization. *Plant Physiol.* 108:1345-1352.
50. Krinsky, S., and R. Wrubel. , 1996. Agricultural biotechnology: Science, policy, and social issues. Univ. Ill. Press, Urbana.
51. Kusaba, S., M. Fukumoto, C. Honda, I. Yamaguchi, T. Sakamoto, and Y. Kano-Murakami. , 1998. Decreased GA(1) content caused by the overexpression of *OSH1* is accompanied by suppression of GA₂₀ and oxidase gene expression. *Plant Physiol.* 117:1179-1184.
52. Landbo, L., B. Andersen, and R.B. Jorgensen., 1996. Natural hybridization between oilseed rape and a wild relative: hybrids among seeds from weedy *B. campestris*. *Hereditas* 125:89-91.
53. Lange, T. , 1998. Molecular biology of gibberellin synthesis. *Planta* 204:409-419.
54. Lange, T., S. Robatzek, and A. Frisse. , 1997. Cloning and expression of gibberellin 2 β ,3 β -hydroxylase cDNA from pumpkin endosperm. *Plant Cell* 9:1459-1467.
55. Lefol, E., V. Danielou, and H. Darmency. , 1996a. Predicting hybridization between transgenic oilseed rape and wild mustard. *Field Crops Res.* 45: 153-161.
56. Lefol, E., A. Fleury, and H. Darmency. , 1996b. Gene dispersal from transgenic crops II. Hybridization between oilseed rape and the wild hoary mustard. *Sex. Plant Reprod.* 9:189-196.
57. Levy, A., 1985. A shattering-resistant mutant of *Papaver bracteatum* Lindl: characterization and inheritance. *Euphytica* 34: 811-815.
58. Li, B. L., and M.E. Foley. , 1997. Genetic and molecular control of seed dormancy. *Trends Plant Sci.* 2:384-389.
59. Lin, S. Y., T. Sasaki, M. Yano. , 1998. Mapping quantitative trait loci controlling seed dormancy and heading date in rice, *Oryza sativa* L., using backcross inbred lines. *Theor. Appl. Genet.* 96:997-1003.
60. Lincoln, J. and Fischer, R., 1988. Diverse mechanisms for the regulation of ethylene-inducible gene expression. *Mol. Gen. Genet.* 212:71-75.

0988977-072004

61. Linder, C.R., 1998. Potential persistence of transgenes: seed performance of transgenic canola and wild X canola hybrids. *Ecol. Appli.* 8:1180-1195.
62. Ling-Hwa, T., and H. Morishima. , 1997. Genetic characterization of weedy rices and the inference on their origins. *Breeding Sci.* 47:153-160
63. Love, S.L. , 1994. Ecological risk of growing transgenic potatoes in the United States and Canada. *Amer. Potato J.* 71:647-658.
64. Lundberg, S., P. Nilsson, and T. Fagerstrom. , 1996. Seed dormancy and frequency dependent selection due to sib competition: The effect of age specific gene expression. *J. Theor. Biol.* 183:9-17.
65. Lutman, P.J.W. , 1993. The occurrence and persistence of volunteer rapeseed (*Brassica napus*). *Asp. Appl. Biol.* 35:29-36.
66. Metz, P.L.J., E. Jacobsen, J.P. Nap, A. Pereira, and W.J. Steikema. , 1997. The impact of biosafety on the phosphinothricin-tolerance transgene in interspecific *B. rapa x B. napus* hybrids and their successive backcrosses. *Theor. Appl. Gen.* 95:442-450.
67. Mikkelsen, T.R., J. Jensen, and R.B. Jorgensen., 1996. Inheritance of oilseed rape (*Brassica napus*) RAPD markers in a backcross with progeny with *Brassica campestris*. *Theor. Appl. Genet.* 92:492-497.
68. Nishijima, T., N. Katsura, M. Koshioka, H. Yamazaki, M. Nakayama, H. Yamane, I. Yamaguchi, T. Yokota, N. Murofushi, N. Takahashi, and M. Nonaka., 1998. Effects of gibberellins and gibberellin-biosynthesis inhibitors on stem elongation and flowering of *Raphanus sativus* L. *J. Jap. Soc. Hort. Sci.* 67:325-330.
69. Pantone, D.J., and J.B. Baker. , 1991. Weed-crop competition models and response-surface analysis of red rice competition in cultivated rice: A review. *Crop Sci.* 31:1105-1110.
70. Paterson, A.H., K.F. Schertz, Y-R. Lin, S-C. Liu and Y-L. Chang., 1995. The weediness of wild plants: Molecular analysis of genes influencing dispersal and persistence of johnsongrass, *Sorghum halepense* (L.) Pers. *Proc. Natl. Acad. Sci. U.S.A.* 92: 6127-6131.
71. Powell, M. , 1997 Science in sanitary and phytosanitary dispute resolution. Discussion Paper 97-50, Resources for the Future, Washington, D.C.
72. Prakash, S. , 1988. Introgression of resistance to shattering in *Brassica napus* from *Brassica juncea* through non-homologous recombination. *Z. Pflanzenzuch.* 101:167-168.

0988977-072004

73. Price, J.S, R.N. Hobson, M.A. Nealle, and D.M. Bruce. , 1996. Seed losses in commercial harvesting of oilseed rape. J. Agr. Engineer. Res. 65:183-191.
74. Regal, P.J., 1994. Scientific principles for ecologically based risk assessment of transgenic organisms. Molec. Ecol. 3:5-13.
75. Rissler, J. and M. Mellon., 1993. Perils amidst the promise - ecological risks of BD-HRCs in a global market. Union of Concerned Scientists, Cambridge MA.
76. Robson, P.R.H., A.C. McCormac, A.S. Irvine, and H. Smith. , 1996. Genetic engineering of harvest index in tobacco through overexpression of a phytochrome gene. Nature Biotech. 14:995-998.
77. Schaller, H., P. Bouvier-Naveo and P. Benveniste., 1998. Overexpression of an *Arabidopsis* cDNA encoding a sterol-C24-methyltransferase in tobacco modifies the ratio of 24-methyl cholesterol to sitosterol and is associated with growth reduction. Plant Physiol. 118:461-469.
78. Scheffler, J.A., R. Parkinson, and P.J. Dale., 1995. Evaluating the effectiveness of isolation distances for field plots of oilseed rape (*Brassica napus*) using a herbicide resistant transgene as a selectable marker. Plant Breed. 114: 317-321.
79. Simon, U., 1994. "Alko" the first seed-shattering resistant cultivar of meadow foxtail *Alopecurus pratensis* L. Acta Hort. 355: 143-146.
80. Sindel, B.M., 1997. Outcrossing of transgenes to weedy relatives. p. 43-81. In: G.D. McLean, P.M. Waterhouse G. Evans and M.J. Gibbs (eds.), Commercialisation of BD-HRCs: Risk, benefit and trade considerations. Coop. Res. Center for Plant Sci. and Bur. of Resource Sci., Canberra.
81. Smith, H., and G.C. Whitelam., 1997. The shade avoidance syndrome: Multiple responses mediated by multiple phytochromes. Plant Cell Environ. 20: 840-844.
82. Smith, M.W., S. Yamaguchi, T. Ait-Ali, and Y. Kamiya. , 1998. The first step of gibberellin biosynthesis in pumpkin is catalyzed by at least two copalyl diphosphate synthases encoded by differentially regulated genes. Plant Physiol. 118: 1411-1419.
83. Snow, A. A., P. Moran-Palma, L.H. Rieseberg, A. Wszelaki, and G.J. Seiler., 1998. Fecundity, phenology, and seed dormancy of F₁ wild-crop hybrids in sunflower (*Helianthus annuus*, Asteraceae). Amer. Jour. Bot. 85:794-801

84. Steber, C. M., S.E. Cooney, P. McCourt, , 1998. Isolation of the GA-response mutant *sly1* as a suppressor of *ABI1-1* in *Arabidopsis thaliana*.. Genetics 149:509-521
85. Thill, D.C., and C.A. Mallory-Smith. , 1997. The nature and consequence of weed spread in cropping systems. Weed. Sci. 45:337-342.
86. Timmons, A.M., Y.M. Charters, J.W. Crawford, D. Burn, S.E. Scott, S.J. Dubbels, N.J. Wilson, A. Robertson, E.T. O'Brien, G.R. Squire and M.J. Wilkinson. 1. Risks from BD-HRCs. Nature 380: 487.
87. Torgersen, H., 1996. Risk assessment in transgenic plants: what can we learn from the ecological impacts of traditional crops? BINAS News 2: (3&4) (<http://www.binas.unido.org/binas/News/96issue34/risk.html>)
88. Torii, K.U., T.W. McNellis, and X.-W. Deng., 1998. Functional dissection of *Arabidopsis* COP1 reveals specific roles of its three structural modules in light control of seedling development. EMBO J. 17:5577-5587.
89. Turner, C.E., 1988. Ecology of invasions by weeds. Weed Management in Agroecosystems: Ecological Approaches. pp. 41-54. In: M.A. Altieri and M. Liebman (eds.), Weed Management in Agroecosystems: Ecological Approaches. CRC Press, Inc. Boca Raton, Florida.
90. N. , 1935. Genome analysis in *Brassica* with special reference to the experimental formation of *B. napus* and the peculiar mode of fertilization. Japan. J. Bot. 7: 389-452.
91. Van der Schaar, W., C. L. A. Blanco, K.M. Kloosterziel, R.C. Jansen, J. W. Van Ooijen, and M. Koornneef. , 1997. QTL analysis of seed dormancy in *Arabidopsis* using recombinant inbred lines and MQM mapping. Heredity 79:, 190-200.
92. Vleeshouwers, L. M. , 1988. The effect of seed dormancy on percentage and rate of germination in *Polygonum persicaria*, and its relevance for crop-weed interaction. Ann. Appl. Biol. 132:289-299.
93. Wan, J., T. Nakazaki, K. Kawaura, and H. Ikehashi., 1997. Identification of marker loci for seed dormancy in rice (*Oryza sativa* L.) Crop Sci. 37: 1759-1763.
94. Waters, S. , 1996. The regulation of herbicide-resistant crops in Europe. p. 347-362. In: S.O. Duke (ed.), Herbicide resistant crops:

P00220 226859

- Agricultural, environmental, economic, regulatory, and technical aspects. CRC Press, Boca Raton.
95. Webb, S. E., N.E.J. Appleford, P. Gaskin, and J. R. Lenton., 1998. Gibberellins in internodes and ears of wheat containing different dwarfing alleles *Phytochemistry* 47:671-677.
 96. White, A.D., M.K.E. Owen, and R.G. Hartzler. , 1998. Evaluation of common sunflower (*Helianthus annuus* L.) resistance to acetolactate synthase inhibiting herbicides. *Weed Sci. Soc. Am. Abstr.* 38:1120.
 97. Whitton, J., D.E. Wolf, D.M. Arias, A.A. Snow, and L.H. Reiseberg. , 1995. The persistence of cultivar alleles in wild populations of sunflowers five generations after hybridization. *Theor. Appl. Genet.* 95:35-40
 98. Williams, M.E. , 1995. Genetic engineering for pollen control. *Trends Biotech. (TIBTECH)* 13:344-349.
 99. Williamson, M., 1993. Invaders, weeds, and risks from genetically manipulated organisms. *Experientia* 49:219-224.
 100. Yamaguchi, S., T.P. Sun, H. Kawaide, and Y. Kamiya. , 1998. The GA₂ locus of *Arabidopsis thaliana* encodes ent-kaurene synthase of gibberellin biosynthesis. *Plant Physiol.* 116:1271-1278.
 101. Young, B.A., 1991. Heritability of resistance to seed shattering in kleingrass., 1991. *Crop Sci.* 31:1156-1158.
 102. Zemetra, R.S., J. Hansen, and C.A. Mallory-Smith. , 1998. Potential for gene transfer between wheat (*Triticum aestivum*) and jointed goatgrass (*Aegilops cylindrica*). *Weed Sci.* 46: 313-317.

WHAT IS CLAIMED IS:

1. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of producing apomictic seeds of said crop of a type which give rise to male sterile crop plants to thereby prevent introgression of the genetically engineered genetic trait of said crop to said weed and to reduce the weedy potential of the crop.

2. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop having multiple genomes derived from different wild sources to a weed having a genome compatible with one of said multiple genomes, the method comprising the step of cytogenetically selecting for genetically engineered crop plants in which a gene or genes responsible for the genetic trait are localized on one or more of said multiple genomes of said crop which is not, or is far less compatible with said genome of said weed.

3. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of controlling the expression of the genetically engineered genetic trait in the crop by at least one control element which is inexpressible by the weed.

4. A genetic construct for genetically modifying a crop to express a genetically engineered genetic trait while mitigating the effects of introgression of the genetically engineered genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment encoding for said genetic trait and at least one additional nucleic acid segment including at least one control element which is expressible by the crop, yet which is inexpressible by the weed.

5. A method of mitigating the effects of introgression of a genetically engineered first genetic trait of a crop to a weed and of mitigating a weedy potential of the crop, the method comprising the step of co-engineering at least one copy of a genetically linked second genetic trait

in said crop, said second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

6. A genetic construct for genetically modifying a crop to express a genetically engineered first genetic trait while mitigating the effects of introgression of the genetically engineered first genetic trait of the crop to a weed, the genetic construct comprising a first nucleic acid segment encoding for said first genetic trait and at least one additional nucleic acid segment encoding a second genetic trait, said second genetic trait being innocuous or somewhat valuable to the crop yet deleterious to the weed.

7. The method or construct according to claims 5 or 6, respectively, wherein said second genetic trait is of abolished secondary dormancy.

8. The method or construct according to claims 5 or 6, respectively, wherein said second genetic trait is of uniform or delayed ripening.

9. The method or construct according to claims 5 or 6, respectively, wherein said second genetic trait is of anti-shattering.

10. The method or construct according to claims 5 or 6, respectively, wherein said second genetic trait is of dwarfism.

11. The method or construct according to claims 5 or 6, respectively, wherein said second genetic trait is selected from the group consisting of seed stalk bolting, seed coat defects that facilitate uniform germination, root storage promotion, biennial growth and non-flowering.

12. A method of mitigating the effects of introgression of a genetically engineered genetic trait of a crop to a weed, the method comprising the step of cytogenetically selecting for or producing genetically engineered crop plants in which a gene or genes responsible for the genetic trait are genetically linked to an endogenous genetic trait of said crop, said endogenous genetic trait being deleterious to the weed.

0988977-072004

Docket No.
01/22288

Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TRANSGENIC PLANTS

the specification of which

☐ is attached hereto.

☒ was filed on 24 January 2000 as PCT

International Application Number PCT/IL00/00046

and was amended on _____

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) or Section 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Not Claimed

128353
(Number)

ISRAEL
(Country)

03/February/1999
(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

I hereby claim the benefit under 35 U.S.C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112. I acknowledge the duty to disclose to the United States Patent and Trademark Office all the information known to me to be material to patentability as defined in Title 37, C.F.R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

PCT/IL00/00046

24 January 2000

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. *(list name and registration number)*

③ **SOL SHEINBEIN, Registration Number 25,457**
BENJAMIN J. BARISH, Registration Number 17,523
MARTIN MOYNIHAN, Registration Number 40,338

Send Correspondence to: **G.E. EHRLICH (1995) LTD.**
c/o ANTHONY CASTORINA
2001 JEFFERSON DAVIS HIGHWAY
SUITE 207
ARLINGTON, VIRGINIA 22202

Direct Telephone Calls to: *(name and telephone number)*

Anthony Castorina

Tel. No. (703) 415-1581

Fax No. (703) 415-4864

FULL NAME OF SOLE OR FIRST INVENTOR		Jonathan GRESSEL
Sole or first inventor's signature <i>Jonathan Gressel</i>		Date 10 July, 2001
Residence	: 15 Hayarden Street, 76 604 Rehovot, Israel	ILX
Citizenship	: ISRAELI and USA (dual)	
Post Office Address	: 15 Hayarden Street, 76 604 Rehovot, Israel	